

创伤性颅脑损伤患者血清 miR-422a, miR-212-5p 表达水平与病情和预后的相关性研究

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摘要: **目的** 探讨创伤性颅脑损伤患者血清 miR-422a, miR-212-5p 表达与病情和预后的相关性, 分析 miR-422a, miR-212-5p 诊断创伤性脑损伤患者预后的价值。 **方法** 连续性选择 2018 年 4 月~2020 年 6 月山东省菏泽市中医医院收治的 173 例创伤性颅脑损伤患者 (创伤组) 和同期 125 例志愿者 (对照组)。根据格拉斯哥昏迷量表 (GCS) 评分将创伤组患者分为轻度组 (GCS 评分 13~15 分, 57 例), 中度组 (GCS 评分 9~12 分, 63 例) 和重度组 (GCS 评分 3~8 分, 53 例), 追踪临床结局, 根据格拉斯哥预后量表 (GOS) 评分将患者分为预后不良组 (GOS 评分 1~3 分, 62 例) 和预后良好组 (GOS 评分 4~5 分, 111 例)。检测所有受试者血清 miR-422a, miR-212-5p 表达, 比较组间差异, 分析 miR-422a, miR-212-5p 与创伤性颅脑损伤患者预后的关系以及 miR-422a, miR-212-5p 预测患者预后的价值。 **结果** 创伤组血清 miR-422a 表达高于对照组 (3.02 ± 1.02 vs 0.95 ± 0.21), miR-212-5p 表达低于对照组 (1.03 ± 0.28 vs 2.85 ± 0.61), 差异均有统计学意义 ($t=22.340, 34.544$, 均 $P < 0.05$)。重度组血清 miR-422a 表达高于中度组和轻度组 (4.01 ± 0.13 vs $2.92 \pm 0.66, 2.21 \pm 0.12$; $t=11.824, 75.516$, 均 $P < 0.05$), miR-212-5p 表达低于中度组和轻度组 (0.84 ± 0.06 vs $0.97 \pm 0.25, 1.27 \pm 0.04$; $t=3.695, 44.512$, 均 $P < 0.05$), 预后不良组血清 miR-422a 表达高于预后良好组 (4.05 ± 0.11 vs 2.44 ± 0.31 , $t=22.340, P < 0.05$), miR-212-5p 表达低于预后良好组 (0.83 ± 0.06 vs 1.14 ± 0.21 , $t=22.340, P < 0.05$), 差异均有统计学意义。低 GCS 评分 (OR=0.825, 95%CI: 0.721~0.945), 高 miR-422a 表达 (OR=1.394, 95%CI: 1.194~1.627), 低 miR-212-5p 表达 (OR=0.744, 95%CI: 0.667~0.831) 是创伤性颅脑损伤患者预后不良的危险因素 (均 $P < 0.05$)。联合 miR-422a, miR-212-5p 预测创伤性颅脑损伤患者预后不良的曲线下面积为 0.907, 高于单独检测 miR-422a, miR-212-5p 的 0.797, 0.775 ($Z=2.412, 2.561$, 均 $P < 0.05$)。 **结论** miR-422a 过表达和 miR-212-5p 表达缺失与创伤性脑损伤严重程度和不良预后有关, 可作为预后预测的潜在生物学指标。

关键词: 微小核糖核酸-422a; 微小核糖核酸-212-5p; 创伤性脑损伤

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Correlation of miR-422a and miR-212-5p Expression Levels with Disease and Prognosis in Patients with Traumatic Brain Injury

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Abstract: Objective To investigate the correlation between the expression of serum miR-422a and miR-212-5p in patients with traumatic brain injury with the condition and prognosis, and analyze the diagnostic value of miR-422a and miR-212-5p in the prognosis of patients with traumatic brain injury. **Methods** A total of 173 patients with traumatic craniocerebral injury (trauma group) and 125 volunteers (control group) admitted to Heze Hospital of Traditional Chinese Medicine of Shandong Province from April 2018 to June 2020 were successively selected. According to the Glasgow Coma Scale (GCS) score, the patients in the trauma group were divided into mild group (GCS score 13~15, 57 cases), moderate group (GCS score 9~12, 63 cases) and severe group (GCS score 3~8, 53 cases). The clinical outcome was tracked, according to Glasgow Outcome Scale (GOS), the patients were divided into poor prognosis group (GOS score 1~3, 62 cases) and good prognosis group (GOS score 4~5, 111 cases). The expression of serum miR-422a and miR-212-5p of all subjects was detected, and the differences between groups were compared to analyze the relationship between miR-422a and miR-212-5p and the prognosis of patients with traumatic brain injury, as well as the value of miR-422a and miR-212-5p in predicting the prognosis of patients. **Results** The expression of miR-422a in the trauma group was higher than that in the control group (3.02 ± 1.02 vs 0.95 ± 0.21), and the expression of miR-212-5p in the trauma group was lower than that in the control group (1.03 ± 0.28 vs 2.85 ± 0.61), the difference sentences were statistically significant ($t=22.340, 34.544$, all $P < 0.05$). The expression of miR-422a in the severe group was higher than that in the moderate and mild groups (4.01 ± 0.13 vs $2.92 \pm 0.66, 2.21 \pm 0.12$; $t=11.824, 75.516$, all $P < 0.05$),

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and the expression of miR-212-5p was lower than that in the moderate and mild groups (0.84 ± 0.06 vs 0.97 ± 0.25 , 1.27 ± 0.04 ; $t = 3.695$, 44.512 , $P < 0.05$). The expression of miR-422a in the poor prognosis group was higher than that in the good prognosis group (4.05 ± 0.11 vs 2.44 ± 0.31 , $t = 22.340$, $P < 0.05$), and the expression of miR-212-5p was lower than that in the good prognosis group (0.83 ± 0.06 vs 1.14 ± 0.21 , $t = 22.340$, $P < 0.05$), the difference sentences were statistically significant, respectively. Low GCS score (OR=0.825, 95%CI: 0.721~0.945), high miR-422a expression (OR=1.394, 95%CI: 1.194~1.627), low miR-212-5p expression (OR=0.744, 95%CI: 0.667~0.831) was risk factors for poor prognosis in patients with traumatic brain injury ($P < 0.05$). The area under the curve of combined miR-422a and miR-212-5p for predicting poor prognosis in patients with traumatic brain injury was 0.907, which was higher than 0.797 and 0.775 of miR-422a and miR-212-5p alone ($Z=2.412$, 2.561 , all $P < 0.05$). **Conclusion** Overexpression of miR-422a and deletion of miR-212-5p were associated with severity and poor prognosis of traumatic brain injury, and can be used as a potential biological indicator for prognosis prediction.

Keywords: miR-422a; miR-212-5p; traumatic brain injury

颅脑损伤是施加于头部的机械力引起的脑形态改变或神经功能破坏,是创伤患者中死亡率和致残率最高的类型^[1]。微小核糖核酸(micro nucleic acid, miRNAs)广泛参与细胞增殖凋亡、代谢、应激、组织修复等几乎生命体各种生命活动的调节过程,在颅脑损伤以及神经组织修复中也发挥重要调节作用^[2]。现有报道显示 miR-422a 在急性缺血性脑卒中急性期表达上调,亚急性期表达下调^[3],在脑组织缺血再灌注过程中,miR-422a 参与神经细胞存活和凋亡的调节过程^[4]。miR-212-5p 在神经元细胞中表达丰富,参与神经元增长分化、突触可塑性、记忆形成等过程,并维持神经元形态和功能^[5],现有研究显示 miR-212-5p 表达异常与阿尔茨海默病发生有关^[6]。然而 miR-422a, miR-212-5p 在颅脑损伤的报道却十分少见,为此本研究通过设计临床对照试验,检测创伤性颅脑损伤患者血清 miR-422a, miR-212-5p 表达特点,分析其与该疾病病情和预后的相关性,以期临床诊治提供借鉴。

1 材料与方法

1.1 研究对象 本研究已经获得山东省菏泽市中医医院伦理委员会批准,自2018年4月~2020年6月连续性选择菏泽市中医医院收治的173例创伤性颅脑损伤患者(创伤组),均有明确外伤史,受伤至入院时间均不超过24h,急诊颅脑多层螺旋CT提示存在颅内血肿或出血病灶,其中男性103例,女性70例,年龄31~52岁,平均年龄 44.15 ± 6.02 岁,致伤机制:交通事故伤75例,高空坠落伤55例,砸伤43例;脑损伤类型:硬膜外血肿32例,硬膜下血肿51例,脑内血肿47例,蛛网膜下腔出血43例。所有患者入院后均由神经外科医生进行格拉斯哥昏迷量表(Glasgow coma Scale, GCS)评分,根据GCS评分将患者分为轻度组(GCS评分13~15分,57例),中度组(GCS评分9~12分,63例)和重度组(GCS评分3~8分,53例)^[7]。排除标准:①脑卒中、颅内肿瘤、颅内感染;②痴呆、阿尔兹海默病、帕金森;③内分泌疾病、免疫和血液系统疾病。另

选择同时段于我院门诊体检的125例志愿者为对照组,均排除神经系统疾病,近期颅脑外伤或手术史,恶性肿瘤以及心、肺、肾等系统性疾病,男性85例,女性40例,年龄38~60岁,平均年龄 44.62 ± 6.51 岁,创伤组和对照组受试者均知情本研究并签署同意书,组间年龄和性别比较均衡性良好($P > 0.05$)。

1.2 仪器与试剂 TRIzol 试剂(上海朝瑞生物科技有限公司), miRNeasy Kit(德国 Qiagen 公司), TaqMan miRNA 逆转录试剂盒(美国 Applied Biosystems 公司), iTaq Universal SYBR Green 超混合液(美国 BIO-RAD 公司), VeritiPro 实时定量 PCR 仪(美国 Applied Biosystems 公司)。

1.3 方法 血清 miR-422a, miR-212-5p 检测:创伤组患者入院后立即(对照组体检当日)采集肘静脉血3ml注入干燥试管,取血液凝固后的上层液离心(4°C , $3\,000\text{r/min}$, 15min, 半径10cm) -80°C 保存待检。RT-PCR 检测 miR-422a, miR-212-5p 表达: TRIzol 法提取总 RNA, miRNeasy Kit 纯化 RNA, TaqMan miRNA 逆转录试剂盒将吸光度值 $A_{260\text{nm}}/A_{280\text{nm}}$ 的 50 ng RNA 样品转录为 cDNA, 加入 iTaq Universal SYBR Green 超混合液,采用实时定量 PCR 仪通过 $2^{-\Delta\Delta C_t}$ 方法 [$2^{-(C_{\text{靶基因}} - C_{\text{参考基因}})}$] 进行 qPCR 定量测量。反应体系 SYBR® Premix Ex Taq™ II ($2 \times$) $12.5\mu\text{l}$, dNTP $1.6\mu\text{l}$, Taq DNA 聚合酶 $1\mu\text{l}$, 上下游引物 $10\mu\text{mol/L}$ 各 $1\mu\text{l}$, 加反应缓冲液至 $20\mu\text{l}$ 。反应条件: 95°C 变性 10s, 65°C 退火 20s; 75°C 延伸 15s, 做 40 个循环。引物序列(由上海基康公司完成): miR-422a 上游 5'-ACTGGACTTAGGGTCAG-3'。下游 5'-GAACATGTCTGCGTATCTC-3'; miR-212-5p 上游 5'-ACCTTGGCTCTAGACTGCT-3', 下游 5'-GCAGGGTCCGAGGTATTC-3'。β-actin(内参)上游: 5'-TGTCCACCTTCCAGCAGATGT-3', 下游: 5'-GCTCAGTAACAGTCCGCCTAGA-3'。做 3 次平行试验, $2^{-\Delta\Delta C_t}$ 计算 miR-422a 和 miR-212-5p 相对表达量,取 3 次平均值。

临床结局追踪:所有患者入院后立即给予心

电监护、颅内压监测、亚低温治疗、脱水降颅压、营养神经、预防脑血管痉挛、维持水电解质平衡等常规治疗,通气障碍者行无创或有创机械通气、意识障碍加重、瞳孔变化者,立即复查颅脑CT,提示血肿量进行性增加达手术指征者立即急诊手术治疗。伤后30天采用格拉斯哥预后量表(Glasgow outcome scale, GOS)^[8]评估颅脑损伤患者预后情况,1分为死亡;2分为植物生存,仅有最小反应;3分为严重残疾,无自我生活能力;4分为轻度残疾,可独立生活;5分为基本恢复良好,遗留轻度缺陷。根据GOS评分将患者分为预后不良组(GOS评分1~3分,62例),预后良好组(GOS评分4~5分,111例)。

1.4 统计学分析 采用SPSS 25.0进行数据分析,计量资料以均数 \pm 标准差($\bar{x}\pm s$)表示,采用单因素方差分析(两两对比采用LSD- t 检验)或独立样本 t 检验,性别以例(n)表示, χ^2 检验差异性。二元Logistic逐步回归分析创伤性脑损伤患者预后不良的危险因素,绘制受试者工作特征曲线(ROC)分析

表1 不同病情亚组血清 miR-422a, miR-212-5p 表达差异 ($\bar{x}\pm s$, $2^{-\Delta\Delta Ct}$)

项目	轻度组 ⁽³⁾ ($n=57$)	中度组 ⁽²⁾ ($n=63$)	重度组 ⁽¹⁾ ($n=53$)	F 值	P 值	(1) vs (2)		(1) vs (3)		(2) vs (3)	
						t	P	t	P	t	P
miR-422a	2.21 \pm 0.12	2.92 \pm 0.66	4.01 \pm 0.13	266.541	0.000	11.824	<0.05	75.516	<0.05	8.000	<0.05
miR-212-5p	1.27 \pm 0.04	0.97 \pm 0.25	0.84 \pm 0.06	111.028	0.000	3.695	<0.05	44.512	<0.05	8.953	<0.05

2.3 不同预后患者血清 miR-422a, miR-212-5p 表达比较 预后不良组血清 miR-422a 表达高于预后良好组(4.05 \pm 0.11 vs 2.44 \pm 0.31),差异具有统计学意义($t=39.486$, $P<0.05$), miR-212-5p 表达低于预后良好组(0.83 \pm 0.06 vs 1.14 \pm 0.21),差异具有统计学意义($t=11.354$, $P<0.05$)。

2.4 miR-422a, miR-212-5p 与创伤性颅脑损伤患者预后的关系 见表2。将年龄、性别(赋值:0=女,1=男),致伤机制(赋值:1=交通事故伤,2=高空坠落伤,3=砸伤),脑损伤类型(赋值:1=硬膜外血肿,2=硬膜下血肿,3=脑内血肿,4=蛛网膜下腔出血),GCS评分,miR-422a, miR-212-5p 纳入 Logistic 回归方程,向后逐步法排除无关变量($\alpha_{进}=0.05$, $\alpha_{出}=0.10$),结果显示低 GCS 评分,高 miR-422a 表达,低 miR-212-5p 表达是创伤性颅脑损伤患者预后不良的危险因素($P<0.05$)。

表2 影响创伤性颅脑损伤患者预后的 Logistic 回归方程

类别	β	SE	Wald χ^2	OR(95%CI)	P 值
GCS评分	-0.192	0.069	7.743	0.825(0.721~0.945)	0.009
miR-422a	0.332	0.079	17.661	1.394(1.194~1.627)	0.000
miR-212-5p	-0.295	0.056	27.750	0.744(0.667~0.831)	0.000

2.5 miR-422a, miR-212-5p 预测创伤性颅脑损伤患者预后不良的价值分析 见图1。miR-422a, miR-

miR-422a, miR-212-5p 诊断创伤性脑损伤患者预后的价值,以 Z 检验曲线下面积的差异性。检验水准 $\alpha=0.05$ 。

2 结果

2.1 创伤组、对照组血清 miR-422a, miR-212-5p 表达比较 见表1。创伤组血清 miR-422a 表达高于对照组(3.02 \pm 1.02 vs 0.95 \pm 0.21),差异具有统计学意义($t=223.340$, $P<0.05$), miR-212-5p 表达低于对照组(1.03 \pm 0.28 vs 2.85 \pm 0.61),差异具有统计学意义($t=35.544$, $P<0.05$)。

2.2 不同病情患者血清 miR-422a, miR-212-5p 表达比较 见表1。重度组血清 miR-422a 表达高于中度组和轻度组,差异具有统计学意义(均 $P<0.05$), miR-212-5p 表达低于中度组和轻度组,差异具有统计学意义(均 $P<0.05$),中度组血清 miR-422a 表达高于轻度组,差异具有统计学意义($P<0.05$), miR-212-5p 表达低于轻度组,差异具有统计学意义($P<0.05$)。

212-5p 预测创伤性颅脑损伤患者预后不良的界值为3.21,0.98,曲线下面积为0.797(95%CI: 0.731~0.863), $P=0.000$,敏感度70.97%,特异度72.07%;0.775(95%CI: 0.700~0.850), $P=0.000$,敏感度72.58%,特异度76.58%。基于二元 Logistic 计算联合 miR-422a, miR-212-5p 的预测值,其曲线下面积为0.907(95%CI: 0.854~0.960), $P=0.000$,敏感度90.32%,特异度91.89%,高于单独 miR-422a, miR-212-5p 预测,差异具有统计学意义($Z=2.412$, 2.561, $P=0.013$, 0.010)。

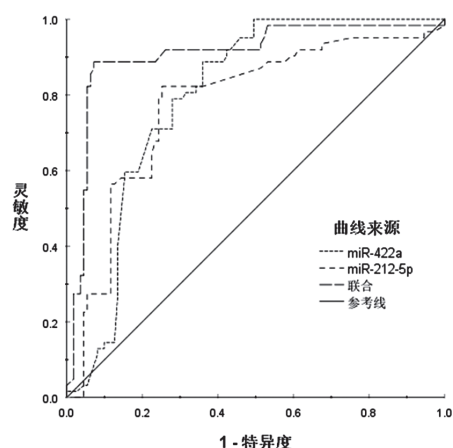


图1 miR-422a, miR-212-5p 预测创伤性颅脑损伤患者预后不良的 ROC 图

3 讨论

miRNA 是受广泛关注的新型小分子非编码 RNA,可调节多达 30% 的蛋白质编码基因的表达,并可能在许多复杂疾病的发展中发挥关键作用^[9]。现有研究显示多种 miRNA 可通过靶向神经病理生理途径参与颅脑外伤过程^[10]。本研究发现 miR-422a 在创伤性脑损伤患者外周血清中表达上调, YAN 等^[11]人检测了创伤性脑损伤血清 miRNA 谱,发现 miR-422a 在轻度颅脑损伤以及严重颅脑损伤中均表达上调,提示 miR-422a 可能与创伤性脑损伤病有关。miR-422a 是作用广泛的 miRNA,定位于 15q22.31,在脑组织中也有广泛表达,现有研究显示氧葡萄糖剥夺/再充氧可诱导脑组织中 miR-422a 表达上调,并诱导神经元细胞毒性,介导神经元细胞凋亡,导致神经元细胞损伤^[12], miR-422a 在免疫介导的脱髓鞘和神经退行性疾病中也出现高表达^[9], miR-422a 可作为缺血性脑卒中的生物学标志物^[3]。miR-422a 参与脑损伤的机制尚不清楚,可能为 miR-422a 可靶向抑制有丝分裂原激活的蛋白激酶 6^[13]和肌细胞增强因子 2D (myocyte enhancement factor 2D, MEF2D)^[14]的表达,其中 MEF2D 是神经元存活所需要的关键转录因子^[15],由此可见 miR-422a 过表达可能促使神经元细胞凋亡,引起病情进展。

本研究结果表明 miR-212-5p 低表达与脑损伤所致昏迷程度加重以及伤后 30 天神经预后不良均有关。miR-212-5p 属于 miR-212 家族成员, miR-212 位于染色体 17p13.3,在中枢神经系统广泛表达,参与神经元形态和功能维持,调控突触活性和认知相关基因表达^[16]。miR-212 表达失调可导致认知相关信号传导通路异常,损害记忆功能^[17], miR-212 还可通过靶向脑微血管内皮细胞中紧密连接蛋白 claudin-1,连接黏附分子 3 和紧密连接相关蛋白 1 表达破坏血脑屏障完整性^[18]。推测 miR-212-5p 参与创伤性脑损伤的机制为:创伤性脑损伤后缺血缺氧可诱导脑组织氧化应激反应,活性氧大量产生,抗氧化物活性降低,继而上调铁中毒相关基因表达,导致铁代谢异常,出现铁累积,最终诱导神经元变性坏死^[19]。miR-212-5p 可靶向环加氧酶-2 (cyclooxygenase-2, COX-2) (又叫 PTGS2), PTGS2 是前列腺素合成的关键酶,可催化脂质氧化,诱导大量脂质活性氧异常累积,导致铁死亡 (Ferroptosis)^[20], Ferroptosis 是一种铁依赖性脂质过氧化物驱使下的非细胞凋亡性死亡方式,颅脑外伤后可诱导 Ferroptosis 产生^[21],因此推测 miR-212-5p 可能通过 miR-212-5p/PTGS2 途径诱导 Ferroptosis,继而导致神经细胞凋亡,神经功能缺损和不良结局发生。

ROC 分析结果显示 miR-422a, miR-212-5p 预测创伤性脑损伤患者预后不良的曲线下面积分别为 0.797, 0.775,而联合两项指标时曲线下面积明显扩大 (0.907),提示联合检测创伤性脑损伤患者伤后 24h 内血清 miR-422a, miR-212-5p 表达水平更有助于判断患者病情和预后,为临床提供更可靠信息。

综上所述,创伤性脑损伤患者血清 miR-422a 表达上调, miR-212-5p 表达下调, miR-422a 过表达和 miR-212-5p 表达缺失与脑损伤严重程度和不良预后有关,可作为预后预测的潜在生物学指标。本研究未观察 miR-422a, miR-212-5p 动态变化,两指标是否与病情变化和临床治疗反应有关尚不清楚,还需进一步研究证实。

参考文献:

- [1] KHELLAF A, KHAN D Z, HELMY A. Recent advances in traumatic brain injury[J]. Journal of Neurology, 2019, 266(11): 2878-2889.
- [2] 樊春荔,何燕娟. 缺血性脑卒中和短暂性脑缺血发作患者血清 miR-23b-3p 水平变化的比较研究[J]. 现代检验医学杂志, 2020, 35(5): 51-54, 123. FAN Chunli, HE Yanjuan. Altered levels of serum miR-23b-3p in patients with ischemic stroke and transient ischemic attack [J]. Journal of Modern Laboratory Medicine, 2020, 35(5): 51-54, 123.
- [3] LI Dongbin, LIU Jingli, WEI Wang, et al. Plasma exosomal miR-422a and miR-125b-2-3p serve as biomarkers for ischemic stroke[J]. Current Neurovascular Research, 2018, 14(4): 330-337.
- [4] DI Yu, LEI Yang, YU Feng, et al. MicroRNAs expression and function in cerebral ischemia reperfusion injury[J]. Journal of Molecular Neuroscience, 2014, 53(2): 242-250.
- [5] WANET A, TACHENY A, ARNOULD T, et al. miR-212/132 expression and functions: within and beyond the neuronal compartment[J]. Nucleic Acids Research, 2012, 40(11): 4742-4753.
- [6] WANG Wangxia, HUANG Qingwei, HU Yanling, et al. Patterns of microRNA expression in normal and early Alzheimer's disease human temporal cortex: white matter versus gray matter[J]. Acta Neuropathologica, 2011, 121(2): 193-205.
- [7] JONES C. Glasgow coma scale[J]. American Journal of Nursing, 1979, 79(9): 1551-1553.
- [8] WARD FULLER G, HERNANDEZ M, PALLOT D, et al. Health state preference weights for the Glasgow outcome scale following traumatic brain injury: a systematic review and mapping study[J]. Value in Health, 2017, 20(1): 141-151.
- [9] SIEGEL S R, MACKENZIE J, CHAPLIN G, et al. Circulating microRNAs involved in multiple sclerosis[J]. Molecular Biology Reports, 2012, 39(5): 6219-6225.
- [10] HUANG Shan, GE Xintong, YU Jinwen, et al. Increased miR-124-3p in microglial exosomes following traumatic brain injury inhibits neuronal

- inflammation and contributes to neurite outgrowth via their transfer into neurons[J]. *FASEB Journal*, 2018, 32(1): 512-528.
- [11] YAN Jing, BU Xiaomin, LI Zhuoling, et al. Screening the expression of several miRNAs from TaqMan Low Density Array in traumatic brain injury: miR-219a-5p regulates neuronal apoptosis by modulating CCNA2 and CACUL1[J]. *Journal of Neurochemistry*, 2019, 150(2): 202-217.
- [12] XU Shu, LI Ya, CHEN Juping, et al. Oxygen glucose deprivation/re-oxygenation-induced neuronal cell death is associated with Lnc-D63785 m6A methylation and miR-422a accumulation[J]. *Cell Death & Disease*, 2020, 11(9): 127-128
- [13] LI Peng, LI Qingmin, ZHANG Yanqiang, et al. MiR-422a targets MAPKK6 and regulates cell growth and apoptosis in colorectal cancer cells[J]. *Biomedicine & Pharmacotherapy*, 2018, 104: 832-840.
- [14] ZHOU Zhixia, LIN Zhijuan, HE Yuqi, et al. The long noncoding RNA D63785 regulates chemotherapy sensitivity in human gastric cancer by targeting miR-422a[J]. *Molecular Therapy-Nucleic Acids*, 2018, 12: 405-419.
- [15] PAZYRA-MURPHY M F, HANS A, COURCHESNE S L, et al. A retrograde neuronal survival response: target-derived neurotrophins regulate MEF2D and bcl-w[J]. *the Journal of Neuroscience*, 2009, 29(20): 6700-6709.
- [16] MENDOZA-VIVEROS L, CHIANG C K, ONG J L K, et al. miR-132/212 modulates seasonal adaptation and dendritic morphology of the central circadian clock[J]. *Cell Reports*, 2017, 19(3): 505-520.
- [17] HANSEN K F, SAKAMOTO K, ATEN S, et al. Targeted deletion of miR-132/-212 impairs memory and alters the hippocampal transcriptome[J]. *Learning & Memory* (Cold Spring Harbor, N.Y.), 2016, 23(2): 61-71.
- [18] BUREK M, KÖNIG A, LANG M, et al. Hypoxia-induced microRNA-212/132 alter blood-brain barrier integrity through inhibition of tight Junction-Associated proteins in human and mouse brain microvascular endothelial cells[J]. *Translational Stroke Research*, 2019, 10(6): 672-683.
- [19] XIE Baoshu, WANG Yiqin, LIN Yong, et al. Inhibition of ferroptosis attenuates tissue damage and improves long-term outcomes after traumatic brain injury in mice[J]. *CNS Neuroscience & Therapeutics*, 2019, 25(4): 465-475.
- [20] STOCKWELL B R, FRIEDMANN ANGELI J P, BAYIR H, et al. Ferroptosis: a regulated cell death nexus linking metabolism, redox biology, and disease[J]. *Cell*, 2017, 171(2): 273-285.
- [21] KENNY E M, FIDAN E, YANG Qin, et al. Ferroptosis contributes to neuronal death and functional outcome after traumatic brain injury[J]. *Critical Care Medicine*, 2019, 47(3): 410-418.

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- [14] HU Yiping, ZHANG Tiantian, CHEN Jingqin, et al. Downregulation of hypoxia-inducible factor-1 α by RNA interference alleviates the development of collagen-induced arthritis in rats[J]. *Molecular Therapy - Nucleic Acids*, 2020, 19: 1330-1342.
- [15] DU Xiangnan, YANG Jian, LIU Cuiying, et al. Hypoxia-Inducible factor 1 α and 2 α have beneficial effects in remote ischemic preconditioning against stroke by modulating inflammatory responses in aged rats[J]. *Frontiers in Aging Neuroscience*, 2020, 12:000 54.
- [16] 陈兰羽, 马继征, 刘咏梅, 等. 基于 HIF-1 α 介导的 VEGF mRNA 表达探讨膈下逐瘀汤抗肝纤维化血管新生的机制 [J]. *中草药*, 2019, 50(2): 449-456.
- CHEN Lanyu, MA Jizheng, LIU Yongmei, et al. Mechanisms of Gexia Zhuyu Decoction on anti-angiogenesis of hepatic fibrosis based on regulation of VEGF mRNA expression mediated by HIF-1 α [J]. *Chinese Traditional and Herbal Drugs*, 2019, 50(2): 449-456.
- [17] 宋颖. 西宁地区妊娠期肝内胆汁淤积症胎盘组织缺氧诱导因子 HIF-1 α 表达及其意义 [J]. *饮食保健*, 2018, 5(28): 286-287.
- SONG Jiong. Expression and significance of hypoxia-inducible factor HIF-1 α in placenta tissue of intrahepatic cholestasis of pregnancy in Xining area[J]. *Diet & Health Care*, 2018, 5(28): 286-287.
- [18] 赵雪晴, 陈辰, 陈亚军. 测定 15 种胆汁酸亚型在妊娠期胆汁淤积症与高胆汁酸血症的鉴别诊断 [J]. *现代妇产科进展*, 2019, 28(12): 917-919.
- ZHAO Xueqing, CHEN Chen, CHEN Yajun. Differential diagnosis of 15 subtypes of bile acid in pregnancy cholestasis and hyperbilirubinemia [J]. *Progress in Obstetrics and Gynecology*, 2019, 28(12): 917-919.
- [19] CELIK S, CALISKAN C S, CELIK H, et al. Predictors of adverse perinatal outcomes in intrahepatic cholestasis of pregnancy[J]. *Ginekologia Polska*, 2019, 90(4): 217-222.
- [20] MAISKAR V, SURVE S. Early onset intrahepatic cholestasis of pregnancy: is progesterone supplementation to be blamed for?[J]. *Journal of Obstetrics and Gynaecology of India*, 2019, 69(2): 192-193.
- [21] 陈霄, 邵勇, 胥飏, 等. 妊娠期无症状高胆汁酸血症的临床特点分析 [J]. *重庆医科大学学报*, 2019, 44(8): 1059-1063.
- CHEN Xiao, SHAO Yong, XU Biao, et al. A clinical analysis of asymptomatic hypercholanemia of pregnancy[J]. *Journal of Chongqing Medical University*, 2019, 44(8): 1059-1063.

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