

血清淀粉样蛋白 A 与肿瘤诊断、分期和预后相关性研究进展

刘振亚, 赵慧敏, 王临艳 (甘肃省妇幼保健院, 兰州 730050)

摘要: 目前用于肿瘤诊断和肿瘤分期的方法多具有侵袭性, 且部分肿瘤标志物敏感性较低, 已不能满足临床需求。已有大量的研究表明: 血清淀粉样蛋白 A (serum amyloid A, SAA) 与多种肿瘤的发生、发展密切相关, 有较高的敏感度和特异度, 无侵袭性、操作简单和成本低等优点。SAA 联合其他肿瘤标志物及肿瘤检测方法在肿瘤诊断、病理分期和预后判定中发挥临床价值。该文就近年来 SAA 在多种肿瘤中的研究进展进行综述, 为肿瘤患者个体化精准治疗、提升治疗效果、提高生存质量、提供理论支持。

关键词: 血清淀粉样蛋白 A (SAA); 肿瘤; 生物标志物

中图分类号: R730.43 **文献标识码:** A **文章编号:** 1671-7414 (2023) 03-207-06

doi: 10.3969/j.issn.1671-7414.2023.03.039

Advances on the Correlation between Serum Amyloid A and Cancer Diagnosis, Stage and Prognosis

LIU Zhen-ya, ZHAO Hui-min, WANG Lin-yan

(Maternal and Child Health Hospital of Gansu Province, Lanzhou 730050, China)

Abstract: At present, most of the methods used for tumor diagnosis and tumor staging are invasive, and some tumor markers have low sensitivity and have been unable to meet the clinical needs. A large number of studies have shown that serum amyloid A (SAA) is closely related to the occurrence and development of a variety of tumors, and has a high sensitivity and specificity, and whether invasive, simple operation and low cost, combined with other tumor markers and tumor detection methods play a clinical value in tumor diagnosis, pathological staging and prognosis. This article reviews the research of SAA in a variety of tumors in recent years, providing theoretical support for individualized precise treatment of cancer patients, improving the therapeutic effect, and improving the quality of life.

Keywords: serum amyloid A (SAA); cancer; biomarker

目前肿瘤仍然是一个严重的全球公共健康问题。根据 2020 年版全球癌症统计显示: 2020 年, 全世界大约新发癌症 1 930 万例, 将近 1 000 万例癌症死亡^[1], 全球癌症发病率和死亡率正在迅速增长。肿瘤的发生较隐匿, 缺乏特异性的症状和体征, 发现时多已到中晚期。早期诊断和治疗监测可以改善癌症患者的预后, 然而大多数血清生物标志物对早期癌症患者的诊断的敏感度和特异度不足^[2], 肿瘤急需早期诊断和治疗的新技术。血清淀粉样蛋白 A (serum amyloid A, SAA) 是一种炎症相关标志物, 参与机体的多种炎症反应。近年来发现 SAA 与多种肿瘤发生、浸润、转移、复发和预后相关^[3], 本文将近年来国内外报道的 SAA 与肿瘤的相关性研究进展进行综述。

1 SAA 的相关概述

1.1 SAA 的结构组成 SAA 是血浆淀粉样蛋白 A, 由 SAA1, SAA2, SAA3 和 SAA4 四个不同的基因编码, 在人类染色体 11p15.1, 是一个跨度 150kb 的

片段, 除 SAA3 基因为 3 外显子和 2 内含子结构外, 其余基因均含有 4 外显子和 3 内含子^[4]。SAA1 和 SAA2 基因彼此相距 15~20kb。成熟的 SAA1 和 SAA2 蛋白长度为 104 个氨基酸, 序列之间 90% 以上相同, 可能来源于进化过程中的基因复制。SAA3 位于 SAA4 基因下游 110 Kb 处。KLUVE-BECKERMAN 等^[5]人之前认为它是一个假基因。然而, LARSON 等^[6]人报道了乳腺上皮细胞中 SAA3 基因的局部转录。SAA4 基因在发现 SAA1 和 SAA2 基因几年后被发现, 它位于 SAA2 基因下游 9kb 处。与 SAA1 和 SAA2 的核苷酸序列相比, SAA4 基因在 69 和 70 密码子之间有 8 个额外的密码子。

1.2 SAA 合成代谢 肝脏是产生 SAA 蛋白的主要场所, SAA 蛋白受各种炎症因子刺激的产生, 在急性免疫反应中发挥着重要作用^[7]。产生是由 IL-6, IL-1 β , 肿瘤坏死因子 (TNF) 等炎症分子以信号转导和依赖转录激活 STAT3 通路方式合成的^[8]。研

基金项目: 甘肃省科学技术厅自然科学基金项目 (20JR10RA424)。

作者简介: 刘振亚 (1989-), 女, 硕士, 主管检验师, 研究方向: 临床检验诊断学, E-mail: lzya1989@126.com。

通讯作者: 王临艳 (1974-), 女, 本科, 副主任检验师, 研究方向: 临床检验诊断学, E-mail: wlyan1974@163.com。

究表明,在急性炎症反应中SAA水平可迅速增加1 000倍,当体内炎症消除后明显降低^[9]。除此以外,SAA也被其他组织和细胞表达和分泌,包括滑膜组织、动脉组织、脂肪细胞和肿瘤组织^[10]。SAA与高密度脂蛋白(HDL)有较高亲和力,与HDL形成SAA/HDL复合物参与机体炎症调控。

1.3 常用检测方法 已报道的SAA检测方法有很多种,我国国家药品监督管理局批准上市的SAA检测试剂盒有50多种,大多使用抗原抗体特异性结合的方法,如:放射免疫分析法、放射免疫扩散法、ELISA,免疫比浊测量法、胶乳增强透射比浊法、化学发光免疫分析法和荧光免疫层析法等^[11],临床应用最广泛的是胶乳增强透射比浊法,该法是将SAA抗体包被在胶乳颗粒上,与标本中SAA抗原特异性结合形成交联微粒,使溶液浊度发生变化,透射吸光度值相应改变,利用标准曲线计算标本中SAA的浓度,该方法具有灵敏度高、操作简便、线性范围广、可用于自动化仪器大样本检测^[12]。

2 SAA与肿瘤的相关性研究

2.1 呼吸系统肿瘤

2.1.1 肺癌:研究发现SAA是区分肺癌与正常人的重要生物标志物,尤其是肺鳞状细胞癌^[13]。王君等^[14]人研究发现,TNM分期越晚血清SAA浓度越高;远端转移者患者血清SAA浓度高于非远端转移患者。体外实验表明,肺癌细胞与人单核细胞白血病(THP-1)细胞相互作用后,肺癌细胞被诱导产生SAA,同时从THP-1单核细胞中诱导出基质金属蛋白酶(MMP-9),基质金属蛋白酶促进肺癌细胞转移。在体内动物模型中,过表达的SAA促进肺癌细胞在肺部转移和定植^[15]。SAA蛋白参与了肺癌肿瘤侵袭和转移,且高水平SAA可能是预测肺癌预后不良的生物标志物^{[16][17]}。

2.1.2 鼻咽癌:SAA是鼻咽癌转移特异性血清生物标志物,鼻咽癌有骨、肝和脾等远处转移的患者SAA的表达明显升高,对鼻咽癌的临床诊断和治疗具有重要意义^[18]。蛋白质芯片技术发现:鼻咽癌复发患者SAA蛋白表达较完全缓解患者明显增加^[19]。CHEN等^[20]人研究显示:SAA较高的鼻咽癌患者预后更差。LI等^[21]人报道了SAA联合EBV DNA检测对生存率的预测准确率和鉴别能力优于TNM分期系统。在鼻咽癌细胞系,SAA1敲除会抑制肿瘤形成和血管生成。其中亚型SAA1.5与 $\alpha V\beta 3$ 整合素的结合亲和力较弱,其蛋白在促进细胞黏附并诱导血管内皮细胞生成中发挥主要作用^[22]。

2.2 消化系统肿瘤

2.2.1 食管癌:研究发现食管癌患者SAA水平显著高于健康人群,SAA水平与食管癌TNM分期、

组织学类型、肿瘤分化程度等相关,SAA联合LncRNA ATB和基质金属蛋白酶-1(MMP-1)可作为食管癌发病风险预测的参考指标^[23]。且术前血清SAA水平升高的食管癌患者的疾病进展快且生存期缩短^[24]。有研究发现食管癌放疗患者中SAA水平较高者,近期疗效较差,生存率较低,易并发食管穿孔^[25]。

2.2.2 胃癌:蛋白质组学技术发现,SAA蛋白峰在胃癌患者血清中明显高于健康组,术后该蛋白峰的表达水平显著下降^[26]。胃癌患者血清SAA水平与FIGO分期、淋巴浸润、远处转移相关,FIGO III-IV期患者SAA水平比FIGO I-II期明显升高。胃癌中癌症相关成纤维细胞(CAF)往往具有促进肿瘤发生的能力,SAA1是胃CAF的候选治疗靶点,SAA1上游增强子H3K27ac和H3K4me1乙酰化可能参与SAA1在胃癌CAF中的过表达^[27]。

2.2.3 胰腺癌:与健康对照组和慢性胰腺炎患者相比,胰腺癌患者血清中SAA显著升高,SAA可以鉴别胰腺癌与良性胰腺炎及健康对照。联合使用SAA、结合珠蛋白及CA19-9生物标志物可以提高胰腺癌诊断的准确性^[28]。手术后胰瘘组SAA的水平明显高于非胰瘘组,SAA对胰腺癌术后胰瘘有重要的辅助诊断价值^[29]。在敲低SAA1基因的人胰腺癌细胞PANC-1的研究发现,胰腺癌细胞的迁移、侵袭和肿瘤耐药性可能是通过NF- κ B激活而增强的^[30]。胰腺导管腺癌存在由癌症相关成纤维细胞(CAF)组成的促纤维增生基质,CAF会刺激肿瘤进展^[31]。

2.2.4 肝癌:与肝良性病变患者相比,肝癌患者SAA水平显著升高,肝癌肿瘤体积较大、BCLC分期较晚的患者血清SAA水平明显较高,血清SAA水平是总生存率的独立预后因素^[32]。相反有文献报道:与正常肝脏组织相比,SAA1在肝细胞癌中表达降低,较低的SAA1表达预示总生存期、预后较差。SAA1的表达随着肿瘤分级和分期的增加而降低,且SAA1表达与TP53突变呈负相关。SAA1的下调可能有助于免疫耐受,这可能是增强抗肿瘤免疫的潜在治疗靶点^[33]。

2.2.5 结直肠癌:结直肠癌术前SAA比正常水平高出2 000倍,术后随着临床病程和化疗周期进展逐渐降低,但从未回到正常范围,当癌症复发时SAA增加^[34]。SAA是可靠的检测结肠癌病程进展和复发的标志物。随着结肠癌分期的增加,血清SAA水平升高。且SAA联合中性粒细胞/淋巴细胞比,血小板/淋巴细胞比可以有效地评价其临床分期,弥补了单一标记物对结肠癌敏感性低的缺陷^[35]。在结肠炎相关小鼠模型中,SAA1/2双敲的小鼠模型中,结肠远端细胞因子IL-4,IL-10和

TNF- α 等减少,巨噬细胞浸润较少,肿瘤的发生率较低。SAA 促进炎症相关损伤和肿瘤发生^[36]。

2.3 泌尿生殖系统肿瘤

2.3.1 乳腺癌:乳腺癌患者血清 SAA 水平明显升高,且与乳腺癌临床分期、淋巴结转移情况及分化程度有关^[37],SAA 水平随乳腺癌分期的增加逐渐升高,有淋巴结转移或远处转移的乳腺癌患者 SAA 浓度明显高于无转移的乳腺癌患者^[38]。因此,SAA 可能是乳腺癌分期和转移的良好候选指标。在体外试验中 SAA1 过表达时下调细胞自噬,在 SAA1/2 敲除的细胞,通过调节自噬,促进细胞抵抗凋亡和坏死。因此,SAA 通过调节乳腺癌细胞的自噬,促进肿瘤的发生^[39]。

2.3.2 卵巢癌:随着上皮细胞从良性和交界性腺瘤发展到原发性和转移性腺癌,SAA 蛋白表达逐渐增加。RT-PCR 分析证实,与正常卵巢组织相比,卵巢癌组织中 SAA1 和 SAA4 基因过表达,卵巢癌细胞系 OVCAR-3 中 SAA mRNA 和蛋白表达较强,SAA 在卵巢癌中的表达增强与卵巢癌的发生有关,并可能具有治疗应用价值^[40]。且血清 SAA 水平与晚期 FIGO 分期、组织学亚型、淋巴浸润和远处转移有显著相关性^[41]。TNF 在人卵巢癌细胞系 OVCAR-3 和 SKOV-3 中显著增加 SAA1/2 水平。由于 SAA1 启动子包含 NF- κ B 位点,NF- κ B 位点在调控 TNF 诱导的 SAA1 启动子活性中起关键作用,卵巢癌上皮细胞中 NF- κ B 激活通过抑制癌前细胞的凋亡促进癌症的发生^[42]。

2.3.3 子宫肿瘤:SAA 基因在子宫内膜癌中的表达明显高于正常子宫内膜。子宫内膜癌中 SAA mRNA 的平均拷贝数是正常子宫内膜细胞的 95 倍。高度纯化的原代子宫内膜癌细胞 SAA 表达呈阳性,在体外能够分泌高水平的 SAA,此结果支持假设:子宫内膜癌患者 SAA 不仅是肝脏分泌的蛋白质,也是子宫内膜癌细胞的产物^[43]。子宫内膜癌 II 和 III 级患者 SAA 水平较正常对照组和 I 级患者明显增高。SAA 鉴别子宫内膜癌的敏感度和特异度分别为 68.7%,58.6%,SAA 联合 CEA,CA125 及 HE4(人附睾蛋白 4)诊断子宫内膜癌的敏感度和特异度分别为 84%,61.1%^[44]。

2.3.4 前列腺癌:前列腺癌患者血清中的 IL-6, SAA 和 PSA 水平明显高于前列腺增生组和正常对照组,随着前列腺癌分化程度的降低,SAA 的水平呈现增高趋势^[45]。血清 SAA 水平与前列腺癌临床分期呈正相关。前列腺骨转移组 SAA 水平较无转移组显著升高。SAA 和 IL-6 联合检测对前列腺癌具有较高的诊断价值,可以作为前列腺癌骨转移的早期诊断指标^[46-47]。

2.3.5 肾癌:肾细胞癌患者血清 SAA 水平升高,明显高于健康对照。SAA 升高组的生存率也明显低于正常 SAA 组。通过单因素和多因素分析,血清 SAA 水平是重要且独立的预后因素^[48]。晚期肾癌伴随着 CRP 和 SAA 的升高,且 SAA 比 CRP 更敏感^[49]。SAA 在晚期肾透明细胞癌中增加,与肾透明细胞癌患者生存率呈负相关。因此,SAA 可作为预测肾透明细胞癌预后不良的标记物^[50]。

2.4 骨肉瘤

双向凝胶电泳技术结果显示,在多形性肉瘤(PS)、软骨肉瘤(CS)和骨肉瘤(OS)患者中,SAA 水平在高度转移的 PS 和 OS 患者中显著升高,而在侵袭性较低的 CS 患者中 SAA 的升高不明显^[51]。当使用 Western blot,ELISA 和质谱估计受试者的 SAA 水平时,也观察到类似的结果。PS,OS 和 CS 等肿瘤恶性程度与患者体内 SAA 水平有明显的相关性。利用划痕实验和 transwell 实验发现 SAA 通过 FPRL-1/ERK/ α v β 3 整合素途径调节骨肉瘤细胞的迁移和侵袭^[52]。

2.5 胶质母细胞瘤(GBM)

星形细胞瘤 I-III 级和胶质母细胞瘤(GBM 或 IV 级)患者比较时,实时荧光定量 PCR 显示 SAA1 mRNA 在 GBM 中明显更高;免疫组化分析显示,在 GBM 细胞质中 SAA 呈阳性。SAA1 可能是区分 GBM 和其他亚型胶质瘤的一个独特基因^[53]。SAA1 表达影响细胞的免疫活性,因此 SAA 是 GBM 免疫治疗中 XAV939, TGX-221 和拉帕替尼的药敏指标^[54]。SAA1 下调促进 GBM 细胞凋亡:SAA1 的下调可以抑制丝氨酸/苏氨酸蛋白激酶 B (AKT) 的磷酸化,从而调控 Bcl2 和 Bax 等凋亡相关蛋白的表达,导致 GBM 细胞死亡^[55]。

3 总结和展望

SAA 是目前临床常用的炎性指标,细菌或病毒感染时 SAA 水平明显升高。SAA 也是慢性炎症和肿瘤发生之间的联系,SAA 水平升高促进肿瘤的发生,加速肿瘤的进展和转移,影响肿瘤的预后。SAA 将成为肿瘤筛查及病情监测的候选生物标志物。SAA 可以联合已有的肿瘤标志物 CA199,CEA,CA125,PSA 等在肿瘤诊断、病理分期和预后判定中发挥临床价值。虽然 SAA 在不同肿瘤中的调控机制尚未完全明确,但将来联合基础实验和临床研究,会明确 SAA 在肿瘤诊断、分期和预后等方面的重要价值,SAA 作为一种新型生物标志物将应用于临床,解决临床问题,受益于肿瘤患者。这是国内外学者们努力解决的问题。

参考文献:

[1] SUNG H, FERLAY J, SIEGEL R L, et al. Global

- Cancer Statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries[J]. *CA Cancer Journal Clinical*, 2021,71(3): 209-249.
- [2] SHIMADA H, NOIE T, OHASHI M, et al. Clinical significance of serum tumor markers for gastric cancer: a systematic review of literature by the Task Force of the Japanese Gastric Cancer Association[J]. *Gastric Cancer*, 2014,17(1): 26-33.
- [3] 李福刚, 雷蕾, 石晓强, 等. 血清淀粉样蛋白 A(SAA) 水平检测在临床实验诊断及健康监测中的应用 [J]. *现代检验医学杂志*, 2019,34(3): 1-5.
LI Fugang, LEI Lei, SHI Xiaoqiang, et al. Serum amyloid A (SAA) test in clinical diagnosis and health monitoring[J]. *Journal of Modern Laboratory Medicine*, 2019, 34(3): 1-5.
- [4] DE BUCK M, COUWY M, WANG Jiming, et al. Structure and Expression of different serum amyloid A (SAA) variants and their concentration-dependent functions during host insults[J]. *Current Medicinal Chemistry*, 2016, 23(17): 1725-1755.
- [5] KLUVE-BECKERMAN B, DRUMM M L, BENSON M D. Nonexpression of the human serum amyloid A three (SAA3) gene[J]. *DNA Cell Biol*, 1991, 10(9): 651-661.
- [6] LARSON M A, WEI S H, WEBER A, et al. Induction of human mammary-associated serum amyloid A3 expression by prolactin or lipopolysaccharide[J]. *Biochem Biophys Res Commun*, 2003, 301(4): 1030-1037.
- [7] TAMAMOTO T, OHNO K, GOTO-KOSHINO Y, et al. Serum amyloid A promotes invasion of feline mammary carcinoma cells[J]. *J Vet Med Sci*, 2014,76(8): 1183-1188.
- [8] ALONZI T, MARITANO D R, GORGONI B, et al. Essential role of STAT3 in the control of the acute-phase response as revealed by inducible gene activation in the liver[J]. *Molecular and Cellular Biology*, 2001, 21(5): 1621-1632.
- [9] KUSHNER I. The acute phase response: an overview[J]. *Methods Enzymol*, 1988,163: 373-383.
- [10] REN Peng, SUN Deshun, XIN Dajing, et al. Serum amyloid A promotes osteosarcoma invasion via upregulating $\alpha v \beta 3$ integrin[J]. *Molecular Medicine Reports*, 2014,10(6): 3106-3112.
- [11] ZHANG Yan, ZHANG Jie, SHENG Huiming, et al. Acute phase reactant serum amyloid A in inflammation and other diseases[J]. *Adv Clin Chem*, 2019, 90: 25-80.
- [12] 中国妇幼保健协会临床诊断与实验医学分会, 柯江维, 徐锦, 等. SAA 单独和与 CRP 联合检测在儿童感染性疾病中的应用专家共识 [J]. *检验医学*, 2021,36(7): 685-690.
Clinical Diagnosis and Laboratory Medicine Branch of China Maternal and Child Health Association, KE Jiangwei, XU Jin, et al. Expert consensus on the application of SAA alone and in combination with CRP in infectious diseases in children[J]. *Laboratory Medicine*, 2021, 36(7): 685-690.
- [13] RONG Biaoxue, LIU Hua, GAO Wenlong, et al. Increased serum amyloid A as potential diagnostic marker for lung cancer: a meta-analysis based on nine studies[J]. *BMC Cancer*, 2016,16(1): 836.
- [14] 王君, 张玉凤, 姜芹. 肺癌患者血清淀粉样蛋白 A(SAA) 水平及临床意义 [J]. *临床肺科杂志*, 2018, 23(8): 1516-1519.
WANG Jun, ZHANG Yufeng, JIANG Qin. Serum amyloid A(SAA) level in patients with lung cancer and its clinical significance[J]. *Journal of Clinical Pulmonary Medicine*, 2018, 23(8): 1516-1519.
- [15] SUNG H J, AHN J M, YOON Y H, et al. Identification and validation of SAA as a potential lung cancer biomarker and its involvement in metastatic pathogenesis of lung cancer[J]. *Journal of Proteome Research*, 2011,10(3): 1383-1395.
- [16] LIN Haiyingjie, TAN Guoqiang, LIU Yan, et al. The prognostic value of serum amyloid A in solid tumors: a meta-analysis[J]. *Cancer Cell Int*, 2019,19: 62.
- [17] CHO W C, YIP T T, CHENG W W, et al. Serum amyloid A is elevated in the serum of lung cancer patients with poor prognosis[J]. *Br J Cancer*, 2010,102(12): 1731-1735.
- [18] LIAO Qiulin, ZHAO Liang, CHEN Xiaodong, et al. Serum proteome analysis for profiling protein markers associated with carcinogenesis and lymph node metastasis in nasopharyngeal carcinoma[J]. *Clin Exp Metastasis*, 2008,25(4): 465-476.
- [19] CHO W C, YIP T T, YIP C, et al. Identification of serum amyloid a protein as a potentially useful biomarker to monitor relapse of nasopharyngeal cancer by serum proteomic profiling[J]. *Clin Cancer Res*, 2004,10(1 Pt 1): 43-52.
- [20] CHEN Qiuyan, TANG Qingnan, TANG Linqun, et al. Pretreatment serum amyloid A and c-reactive protein comparing with epstein-barr Virus DNA as prognostic indicators in patients with nasopharyngeal carcinoma: a prospective study[J]. *Cancer Research and Treatment*, 2018,50(3): 701-711.
- [21] LI Jianpei, LAI Changchun, PENG Songguo, et al. The prognostic value of integration of pretreatment serum amyloid A (SAA)-EBV DNA (S - D) grade in patients with nasopharyngeal carcinoma[J]. *Clinical and Translational Medicine*, 2020,9(1): 2.
- [22] LUNG H L, MAN O Y, YEUNG M C, et al. SAA1 polymorphisms are associated with variation in antiangiogenic and tumor-suppressive activities in nasopharyngeal carcinoma[J]. *Oncogene*, 2015,34(7): 878-889.
- [23] 李筱, 张静, 向慧敏. 血浆长链非编码 RNA ATB 联合 MMP-1 与 SAA 在食管癌中的临床价值研究 [J]. *实用癌症杂志*, 2019, 34 (8): 1244-1249, 1260.
LI Xiao, ZHANG Jing, XIANG Huimin. Clinical Value of Long Non-coding RNA ATB combined with MMP-1 and SAA in esophageal carcinoma[J]. *The Practical Journal of Cancer*, 2019,34(8): 1244-1249, 1260.
- [24] WANG Junye, ZHENG Yuzhen, YANG Juan, et al. Elevated levels of serum amyloid A indicate poor

- prognosis in patients with esophageal squamous cell carcinoma[J]. *BMC Cancer*, 2012,12: 365.
- [25] 胡勇,刘亚军,张强,等.血清淀粉样蛋白A对食管癌再程放疗患者预后的影响[J].*中国肿瘤临床与康复*, 2021,28(4): 439-442.
HU Yong, LIU Yajun, ZHANG Qiang, et al. Serum amyloid A and prognosis of esophageal cancer on reirradiation[J].*Chin J Clin Oncol Rehabil*, 2021,28(4): 439-442.
- [26] 刘池波,梁勇,王海宝,等.胃癌患者中血清淀粉样蛋白A的测定及临床意义[J].*中华胃肠外科杂志*, 2009, 12(3): 314-315.
LIU ChiBo, LIANG Yong, WANG Haibao, et al. Determination and clinical significance of serum amyloid albumen A in patients with gastric cancer[J].*Chin J Gastrointest Surg*, 2009,12(3): 314-315.
- [27] YASUKAWA Y, HATTORI N, IIDA N, et al. SAA1 is upregulated in gastric cancer-associated fibroblasts possibly by its enhancer activation[J]. *Carcinogenesis*, 2021,42(2): 180-189.
- [28] FIRPO M A, GAY D Z, GRANGER S R, et al. Improved diagnosis of pancreatic adenocarcinoma using haptoglobin and serum amyloid A in a panel screen[J]. *World J Surg*, 2009,33(4): 716-722.
- [29] 陈国利,羿海钊,王建利.血清PCT, SAA, CRP, ALB对胰腺癌术后胰瘘的诊断价值[J].*河北医学*, 2022,28(5): 853-857.
CHEN Guoli, YI Haizhao, WANG Jianli. Diagnostic value of the change of serum PCT, SAA, CRP and ALB in pancreatic fistula after surgery for pancreatic cancer[J]. *Hebei Medicine*, 2022, 28(5): 853-857.
- [30] TAKEHARA M, SATO Y, KIMURA T, et al. Cancer - associated adipocytes promote pancreatic cancer progression through SAA1 expression[J]. *Cancer Science*, 2020,111(8): 2883-2894.
- [31] DJUREC M, GRANA O, LEE A, et al. SAA3 is a key mediator of the protumorigenic properties of cancer-associated fibroblasts in pancreatic tumors[J]. *Proc Natl Acad Sci U S A*, 2018,115(6): E1147-E1156.
- [32] NI Xiaochun, YI Yong, FU Yipeng, et al. Serum amyloid A is a novel prognostic biomarker in hepatocellular carcinoma[J]. *Asian Pacific Journal of Cancer Prevention*, 2015,15(24): 10713-10718.
- [33] ZHANG Wei, KONG Huifang, GAO Xudong, et al. Immune infiltration-associated serum amyloid A1 predicts favorable prognosis for hepatocellular carcinoma[J]. *World Journal of Gastroenterology*, 2020,26(35): 5287-5301.
- [34] GLOJNARIC I, CASL M T, SIMIC D, et al. Serum amyloid A protein (SAA) in colorectal carcinoma[J]. *Clin Chem Lab Med*, 2001,39(2): 129-133.
- [35] YANG Qinghua, SUN Chengcheng, ZHAO Lisha. Expression and predictive value of serum NLR, PLR combined with SAA in patients with different stages of colorectal cancer[J]. *Frontiers in Surgery*, 2022, 9: 906074.
- [36] DAVIS T A, CONRADIE D, SHRIDAS P, et al. Serum amyloid a promotes inflammation-associated damage and tumorigenesis in a mouse model of colitis-associated cancer[J]. *Cellular and Molecular Gastroenterology and Hepatology*, 2021,12(4): 1329-1341.
- [37] 雷建梅,匡文斌,邓秋婵,等.血清HIF-1 α , 25(OH)D3, IGF-1及SAA水平在不同临床特征乳腺癌患者中的表达及临床意义[J].*中国医学创新*, 2021,18(34): 48-52.
LEI Jianmei, KUANG Wenbin, DENG Qiuchan, et al. Expression and clinical significance of levels of HIF-1 α , 25(OH)D3, IGF-1 and SAA in breast cancer patients with different clinical characteristics[J]. *Medical Innovation of China*, 2021,18(34): 48-52.
- [38] ZHANG Guojun, SUN Xudong, LÜ Hong, et al. Serum amyloid A: A new potential serum marker correlated with the stage of breast cancer[J]. *Oncol Lett*, 2012,3(4): 940-944.
- [39] PLESSIS M, DAVIS T A, OLIVIER D W, et al. A functional role for serum amyloid A in the molecular regulation of autophagy in breast cancer[J]. *Frontiers in Oncology*, 2022,12(10): 925-944.
- [40] URIELI-SHOVAL S, FINCI-YEHESKEL Z, DISHON S, et al. Expression of serum amyloid a in human ovarian epithelial tumors: implication for a role in ovarian tumorigenesis[J]. *Journal of Histochemistry & Cytochemistry*, 2010,58(11): 1015-1023.
- [41] LI Ze, HOU Yongwang, ZHAO Meng, et al. Serum amyloid A, a potential biomarker both in serum and tissue, correlates with ovarian cancer progression[J]. *Journal of Ovarian Research*, 2020,13(1): 67.
- [42] CHOI H, IGNACIO R M C, LEE E, et al. Augmented serum amyloid A1/2 mediated by TNF-induced NF- κ B in human serous ovarian epithelial tumors[J]. *Immune Network*, 2017,17(2): 121.
- [43] COCCO E, BELLONE S, EL-SAHWI K, et al. serum amyloid A (SAA): a novel biomarker for endometrial[J]. *Cancer*, 2010,116(4): 843-851.
- [44] OMER B, GENC S, TAKMAZ O, et al. The diagnostic role of human epididymis protein 4 and serum amyloid-A in early-stage endometrial cancer patients[J]. *Tumor Biology*, 2013,34(5): 2645-2650.
- [45] 陶小枫,王晓希,刘维,等.血清IL-6, SAA及PSA检测对前列腺癌的诊断价值[J].*山西医科大学学报*, 2019,50(01): 70-74.
TAO Xiaofeng, WANG Xiaoxi, LIU Wei, et al. Diagnostic value of serum IL-6, SAA and PSA for prostate cancer[J]. *J Shanxi Med Univ*, 2019,50(1): 70-74.
- [46] 陈维真,张勇,罗辉. SAA与前列腺癌病理分级和临床分期的关系[J].*当代医学*, 2010,16(25): 16-17.
CHEN Weizhen, ZHANG Yong, LUO Hui. Relationship between SAA and pathological grade and clinical stage of prostate cancer[J]. *Contemporary Medicine*, 2010, 16(25): 16-17.
- [47] 程清,杨涛,尹全乐.血清前列腺特异性抗原同源异构体2及淀粉样蛋白A对前列腺癌的诊断价值[J].*山西医药杂志*, 2020,49(19): 2649-2650.
CHEN Qing, YANG Tao, YING Quanle. Diagnostic value of serum prostate-specific antigen homologous

- isomer 2 and amyloid protein a in prostate cancer[J]. Shanxi Med J, 2020, 49(19): 2649-2650.
- [48] KIMURA M, TOMITA Y, IMAI T, et al. Significance of serum amyloid A on the prognosis in patients with renal cell carcinoma[J]. Cancer, 2001, 92(8): 2072-2075.
- [49] FISCHER K, THEIL G, HODA R, et al. Serum amyloid A: a biomarker for renal cancer[J]. Anticancer Res, 2012, 32(5): 1801-1804.
- [50] OZ ATALAY F, AYTAC VURUSKAN B, VURUSKAN H. Significance of amyloid A immunoexpression in the prognosis of renal cell carcinoma[J]. APMIS, 2016, 124(4): 257-262.
- [51] WAN-IBRAHIM W I, ASHRAFZADEH A, SINGH V A, et al. Contrasting increased levels of serum amyloid A in patients with three different bone sarcomas: An indicator of tumor malignancy?[J]. Electrophoresis, 2016, 37(17/18): 2328-2337.
- [52] REN Peng, SUN Deshun, XIN Dajing, et al. Serum amyloid A promotes osteosarcoma invasion via upregulating $\alpha v \beta 3$ integrin[J]. Mol Med Rep, 2014, 10(6): 3106-3112.
- [53] KNEBEL F H, UNO M, GALATRO T F, et al. Serum amyloid A1 is upregulated in human glioblastoma[J]. Journal of Neuro-Oncology, 2017, 132(3): 383-391.
- [54] CAO Kangxi, JIANG Xingyu, WANG Baishun, et al. SAA1 expression as a potential prognostic marker of the tumor microenvironment in glioblastoma[J]. Front Neurol, 2022, 13: 905561.
- [55] ZHANG Huikai, XU Yang, DENG Gang, et al. SAA1 knockdown promotes the apoptosis of glioblastoma cells via downregulation of AKT signaling[J]. J Cancer, 2021, 12(9): 2756-2767.
- 收稿日期: 2022-12-26
修回日期: 2023-04-07

(上接第206页)

- MEI Yan, ZHANG Ping, JIN Minfei, et al. Research of pregnancy associated group B *Streptococcal* infection[J]. Chinese Journal of Perinatal Medicine, 2017, 20(12): 895-898.
- [2] VIELOT N A, TOVAL-RUIZ C E, WEBER R P, et al. Prevention of group B *Streptococcal* early-onset disease in newborns: ACOG committee opinion summary, number 797[J]. Obstetrics and Gynecology, 2020, 135(2): e51-e72.
- [3] GUAN Xiaoshan, MU Xiaoping, JI Wenjing, et al. Epidemiology of invasive group B *Streptococcal* disease in infants from urban area of South China, 2011-2014[J]. BMC Infectious Diseases, 2018, 18(1): 14.
- [4] 刘娜, 尤建萍, 王博, 等. 妊娠晚期孕妇阴道 B 族链球菌感染青霉素钠治疗对血浆凝血功能及新生儿结局的影响分析[J]. 现代检验医学杂志, 2019, 34(4): 146-150.
- LIU Na, YOU Jianping, WANG Bo, et al. Analysis on the influences of penicillin Sodium treatment on plasma coagulation functions and neonatal outcomes in late pregnant women with vaginal group B *Streptococcus* infections[J]. Journal of Modern Laboratory Medicine, 2019, 34(4): 146-150.
- [5] 中华医学会围产医学分会, 中华医学会妇产科学分会产科学组. 预防围产期 B 族链球菌病(中国)专家共识[J]. 中华围产医学杂志, 2021, 24(8): 561-566.
- Society of Perinatal Medicine, Chinese Medical Association, Obstetrics Subgroup, Society of Obstetrics and Gynecology, Chinese Medical Association. Chinese experts consensus on prevention of perinatal group B *Streptococcal* disease [J]. Chinese Journal of Perinatal Medicine, 2021, 24(8): 561-566.
- [6] PANGERL S, SUNDIN D, GERAGHTY S. Group B *Streptococcus* screening guidelines in pregnancy: a critical review of compliance[J]. Maternal and Child Health Journal, 2021, 25(2): 257-267.
- [7] FILKINS L, HAUSER J R, ROBINSON-DUNN B, et al. American society for microbiology provides 2020 guidelines for detection and identification of group B *Streptococcus*[J]. Journal of Clinical Microbiology, 2020, 59(1): e01230-20.
- [8] 曹清芸, 柏明见, 何美琳, 等. B 族链球菌在妊娠末期孕妇中的感染状态与阴道微生态评分相关性分析[J]. 现代检验医学杂志, 2019, 34(2): 122-124.
- CAO Qingyun, BAI Mingjian, HE Meilin, et al. Correlation between Group B *Streptococcal* infection and nugent score at the third trimester of pregnancy[J]. Journal of Modern Laboratory Medicine, 2019, 34(2): 122-124.
- [9] MADRID L, SEALE A C, KOHLI-LYNCH M, et al. Infant group B *Streptococcal* disease incidence and serotypes worldwide: systematic review and meta-analyses[J]. Clinical Infectious Diseases, 2017, 65(suppl 2): S160-S172.
- [10] 肖艳群, 王华梁. 临床分子诊断质量管理问题及思考[J]. 中华检验医学杂志, 2018, 41(2): 85-87.
- XIAO Yanqun, WANG Hualiang. Problems and reflection on quality management of clinical molecular diagnosis[J]. Chinese Journal of Laboratory Medicine, 2018, 41(2): 85-87.
- [11] ALLEN V M, YUDIN M H, Infectious Diseases Committee. Management of group B *Streptococcal* bacteriuria in pregnancy[J]. Journal of Obstetrics and Gynaecology Canada, 2012, 34(5): 482-486.
- [12] 张睿, 徐英春, 吴洁, 等. B 族链球菌核酸检测试剂的性能评价[J]. 中国医学装备, 2019, 16(9): 46-49.
- ZHANG Rui, XU Yingchun, WU Jie, et al. Validation of nucleic acid detection performance of group B *Streptococcus* (GBS)[J]. China medical equipment, 2019, 16(9): 46-49.
- 收稿日期: 2022-09-07
修回日期: 2023-02-09